# BIOCHEMICAL EFFECTS OF SOME ORGANOPHOSPHORUS PESTICIDES ON THE OVARIES OF ALBINO RATS

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Abstract : An evaluation of the toxic effects of three organophosphates; monocrotophos, dimethoate and methyl parathion on female reproduction was made by biochemical estimations of cytoplasmic and membrance bound proteins, lipids, phospholipids and cholesterol in the rat ovaries after treatment with their low residual level doses ( $LD_{50}$  1/8–1/5) to three groups of six rats each for 90 days. All the three pesticides caused degenerative changes in the ovaries as evidenced by a significant decrease in the concentration of cytoplasmic as well as membrane bound proteins, total lipids, phospholipids and cholesterol. The observations are thus indicative of the reproductive toxicity caused by organophosphates at cellular and molecular level in the ovaries of rats.

Key	words	:	organophosphates	rats	ovaries	proteins
			lipids	phospholipic	ls	cholesterol

### INTRODUCTION

Organophosphates are commonly used as insecticides in agriculture in India and are potent toxicants (1). A serious problem with some organophosphorus compounds has been their high acute toxicity to man and non-target organisms (2). Many environmental pollutants are known to cause reproductive toxicity and have resulted in a decrease in the fertility level of human population (3), but a few reproductive toxicants have been clearly identified and the mechanisms of chemical toxicity on reproductive organs are not well defined (4). Therefore, in the present studies, it was planned to investigate the biochemical effects of three organophosphates; dimethoate, methyl parathion and monocrotophos, after their long-term treatment in low residual level doses on ovaries of albino rats.

## METHODS

#### Animals

Adult (1½-2 months age; 150-200 g body weight) female rats of Wistar strain were housed in groups of two per cage, maintained under controlled conditions of temperature ( $22 \pm 2^{\circ}$ C) and light (12:12 L:D) and provided with food and water ad libitum. Cyclicity of rats was confirmed by examining their vaginal smears daily and the ones showing two or more regular 4–5 days estrous cycles were selected for the experiment.

#### Treatment with organophosphates

Groups of six rats each were given daily oral doses of dimethoate  $(1/5 \text{ LD}_{50}; 76 \text{ mg/} \text{kg})$ , methyl parathion  $(1/5 \text{ LD}_{50}; 4.8 \text{ mg/} \text{kg})$  and monocrotophos  $(1/8 \text{ LD}_{50}; 2.9 \text{ mg/} \text{kg})$  for 90 days. An equal number of animals were vehicle (water) treated and kept as control for each pesticide.

#### **Biochemical** estimations

A day after the last dose, the animals were sacrificed by decapitation, ovaries and liver extirpated, put in normal saline and freed from fat and adhering connective tissues. The cleared ovaries and liver were soaked on a piece of filter paper, weighed, minced and homogenized in known volume of ice-cold phosphate - buffer saline (PBS, pH 7.4) and the ovarian homogenate was centrifuged.

#### Determination of proteins

The soluble (cytoplasmic) proteins were estimated in the supernatant and the residue (pellet) was washed with PBS by centrifugation at 1000 rpm for 5 minutes. The supernatant was separated and pellet dissolved in PBS and shaken with known

volume of 1% solution of the detergent deoxycholate (DOC) for 1 hr at 37°C. The solution was centrifuged at 10,000 rpm for 30 minutes, pellet discarded and supernatant used for estimation of membrane bound proteins and lipids. In the liver homogenate and both the components of ovaries, total proteins were estimated by the method of Lowry et al (5) and expressed as mg/g fresh liver/ovary.

# Determination of total lipids, phospholipids and cholesterol

The liver homogenate and both the supernatants ovarian left after the estimation of proteins were used for the estimation of total lipids by the method of Folch et al (6). Total lipids expressed as mg/g fresh liver or ovary were further analysed for phospholipids and cholesterol by using standard techniques of Ames (7) and Zlatkis and Zak (8) respectively and expressed as mg/g of fresh liver/ovary as well as percent of total lipids. The data were subjected to statistical analysis applying student's t-test.

## RESULTS

# Effects of OPs on ovarian proteins, lipids, phospholipids and cholesterol

The concentration of total cytoplasmic as well as membrane bound proteins, lipids, phospholipids and cholesterol significantly decreased in the ovaries of rats treated with each of the organophosphate (Table I and II). The decrease in concentrations of these biochemical components was statistically significant (P<0.05-0.1) when compared with their respective control levels. (Table I and II). The concentration of phospholipids and cholesterol, when expressed in terms of percentage of total lipid content of the

Treatment	Proteins (mg/g)	Total lipids (mg/g)	Phospholipids (mg/g)	Cholestrol (mg/g)
Control	30.49±5.86	44.72±15.52	5.69±1.83	2.65±0.19
Dimethoate	$23.61 \pm 1.50$	$14.03 \pm 16.46$	$2.28 \pm 0.74*$	$1.05 \pm 0.26*$
Control	$32.50 \pm 6.90$	$37.41 \pm 13.50$	$4.95 \pm 1.63$	$2.51 \pm 0.15$
Methyl parathion	$19.19 \pm 5.32*$	$21.05 \pm 4.76$	$1.50 \pm 0.35*$	0.16±0.009**
Control	$28.03 \pm 3.77$	$40.15 \pm 6.49$	$4.93 \pm 0.31$	$2.55 \pm 0.04$
Monocrotophos	$14.87 \pm 1.44*$	9.68±1.44**	$1.90 \pm 1.0*$	$0.44 \pm 0.02 **$

TABLE I: Effect of some organophosphates on concentration of cytoplasmic proteins, total lipid, phospholipids and cholesterol in the ovaries of albino rats (Mean ± SD).

\*P<0.05; \*\*P<0.1

TABLE II: Effect of some organophosphates on membrane bound (DOC extracted) proteins, total lipid, phospholipids and cholesterol in the ovaries of albino rats (Mean ± SD).

Treatment	Proteins (mg/g)	Total lipids (mg/g)	Phospholipids (mg/g)	Cholestrol (mg/g)
Control	$11.79 \pm 1.67$	102.07±36.45	10.46±2.18	2.83±0.96
Dimethoate	$5.14 \pm 1.04 * *$	79.42±6.19**	3.95±0.63**	1.10±0.49**
Control	$2.47 \pm 2.86$	$110.73 \pm 16.48$	$9.06 \pm 1.37$	$2.19 \pm 0.16$
Methyl parathion	$6.05 \pm 0.90 * *$	77.00±4.18**	$7.77 \pm 1.26$	0.93±0.06**
Control	$10.31 \pm 3.27$	$96.00 \pm 11.79$	8.56±11.79	$2.61 \pm 0.11$
Monocrotophos	$3.08 \pm 0.3 * *$	35.96±3.18**	2.87±0.64**	$0.95 \pm 0.12 * *$

\*P<0.1

TABLE III: Effect of some organophosphates on the percentage of cholesterol and phospholipids in total lipids in ovaries of albino rats.

	% in cyt total	oplasmic lipids	% in DOC extracted membrane lipids	
Ireatment -	Choles- terol (%)	Phospho- lipids (%)	Choles terol (%)	Phospho (%) lipids
Control	5.93	1.27	2.77	10.25
Dimethoate	7.48	16.25	1.39	4.97
Control	6.71	13.23	1.98	8.18
Methyl parathior	n 0.76	7.13	1.21	10.09
Control	6.35	12.28	2.72	8.92
Monocrotophos	4.56	19.63	2.64	7.98

ovaries, was also found to be decreased after treatment with all the OPs (Table III). The concentration of proteins and lipids also decreased in the liver of OP treated rats as compared to that in their control rats (Table IV).

TABLE IV: Effect of some organophosphates on protein and lipid content in liver of female albino rats (mean  $\pm$  SD).

Treatment	Protein (mg/g)	Lipid (mg/g)	
Control	7.04±0.13	29.16±4.77	
Dimethoate	$5.97 {\pm} 0.35 {*}$	$20.83 \pm 4.21*$	
Control	$6.47 {\pm} 0.26$	$28.90{\pm}2.50$	
Methyl parathion	$4.73 \pm 0.07*$	$19.66 \pm 4.21*$	
Control	$6.56 {\pm} 0.86$	$33.33{\scriptstyle\pm}4.21$	
Monocrotophos	$4.95 {\pm} 0.18 {*}$	$25.00 \pm 0*$	

\*P<0.05

# DISCUSSION

The decrease in the proteins, lipids, phospholipids and cholesterol (cytoplasmic and membrane bound) in the ovaries of rats after long term treatment with low doses of dimethoate, methyl parathion and monocrotophos indicates degenerative changes. A decrease in the total lipid content in schiatic nerve, kidney, skeletal muscle, brain and spinal cord was also observed after treatment of chicks with organophosphates; two disyston and folithion (9). Phospholipids and the ratio of phospholipids to total cholesterol also decreased in the nervous tissue of the chicks with these two pesticides, which are indicative of degenerative changes (9). Similar lipid changes have also been reported with other organophosphates like lebaycid and metasystox-R (10). The toxicity of organophosphates varies with structures (WHO, 1986). A11 organophosphorus pesticides (OP) are lipophilic and these environmental xenobiotics are known to have a strong affinity for interaction with membrane phospholipids (11)and the phospholipid component of the biomembrane

is believed to be the site of action of OP insecticides (12). There are also evidences that oxygen free radical formation can be a factor in the toxicity of organophosphates (13). One of the targets of creative oxygeninduced injury is lipid peroxidation, and peroxidation of membrane phospholipids not only alters lipid milieu and structural and functional integrity of cell membrane but also affects the activities of various membrane bound enzymes including total ATPase, Na+-K<sup>+</sup>-ATPase and Mg-ATPase (14-16). Thus in addition to acctylcholinestrase being the principal mode of action of OP pesticides (17) increased lipid peroxidation (12-13; 18-20) and altered ATPase (21-26) have also been implicated in mediating OP toxicity in animals. The inhibitory alterations in the membrane bound lipid composition of the OP treated rat ovaries in the present observations might be result of their this activity and present studies are thus indicative of such degenerative changes induced by dimethoate, methyl parathion and monocrotophos at the cellular and molecular level in the ovaries and liver of rats after their long term exposure even in very low residual level doses.

#### REFERENCES

- 1. Jayaraj S. Intensive use of agricultural chemicals in India and their ecological and environmental impacts. Proc Third Agri Sci Cong, Punjab Agricultural University, Ludhiana 1997: 115-130.
- 2. Namba T, Nolte CT, Jackrel G, Grob D. Poisoning due to organophosphate insecticides: acute and chronic manifestations. *Am J Med* 1977; 50: 475-492.
- Whorton D, Krauss RM, Marshall S, Milby TH. Infertility in male pesticide workers. *Lancet* 1977; 2: 1259-1261.
- 4. Lasley BL, Overstreet JW. Biomarkers for

assessing human female reproductive health, an interdisciplinary approach. *Environ Health Persp* 1998; 106: 955–960.

- Lowry OH, Rosenbrough NJ, Fair AL, Randall RJ. Protein measurement with the Folin - Phenol reagent. J Biol Chem 1951; 193: 265-270.
- Folch J, Lees M, Sloane Stanley GH. A simple method for the isolation and purification of total lipids from animal tissues. J Biol Chem 1957; 226: 497-509.
- Ames BN. In: Methods of Enzymology (Eds. Nevfeld EF and Guisbory VJ) Academic Press, New York, Vol. VIII, 1967 256 p.

- 8. Zlatkis A, Zak B. Study of a new Cholesterol reagent. Anal Biochem 1968; 29: 143-48.
- Gopal PK, Ahuja SP. Lipid and growth changes in organs of chicks (Callus domesticus) during acute and chronic toxicity with Disyston Folithion. *Indian* J Exp Biol 1979; 17: 1153-1154.
- Schoental R. Alkylation of coenzymes and the acute effects of alkylating hepatotoxin. FEBS Lett 1976; 61: 1153-1154.
- Antunes Madeira MC, Madeira VMC. Partition of malathion in synthetic and native membranes. *Biochem Biophys Acta* 1987; 901: 61-67.
- Datta C, Dasgupta JG, Sengupta D. Interaction of organophosphorus insecticides phosphamidon and malathion on lipid profile and acetylcholinesterase activity in human erythrocyte membrane. *Indian* J Med Res 1994; 100: 87-94.
- Bagchi D, Bagchi M, Hassoun EA, Stohs SJ. In vitro and in vivo generation of reactive oxygen species, DNA damage and lactate dehydrogenase leakage by selected pesticides. Toxicology 1995; 104: 129-136.
- Shao Q, Matsubara T, Bhatt SK, Dhalla NS. Inhibition of cardiac sarcolemma Na<sup>+</sup>-K<sup>+</sup>- ATPase by oxyradical generating systems. *Mol Cell Biochem* 1995; 147: 139.
- Rauchova H, Ledvinkova J, Kalous M, Drahota Z. The effect of lipid peroxidation on the activity of various membrane-bound ATPases in rat kidney. *Int J Biochem Cell Biol* 1995; 27: 251-261.
- Fritz KI, Groenendaal F, Andersen C, Ohnishi ST, Mishra OP. Deleterious brain cell membrane effects after NMDA receptor antagonist administration to new born piglets. *Brain Res* 1999; 816: 438.
- Kaur S, Dhanju CK. Enzymatic changes induced by some organophosphorus pecticides in female rats. *Indian J Exp Biol* 2004; 42: 1017–1019.

- Mathews MS, Devi K.S. Effect of chronic exposure of pregnant rats to malathion and/or estrogen and/or progesterone on xenobiotic metabolizing enzymes. *Pest Blochem Physiol* 1994; 48: 110-120.
- Yamano T, Morita S. Effects of pesticides on isolated rat hepatocytes, mitochondria and microsomes II. Arch Environ Contain Toxicol 1995; 28: 1.
- Yang ZP, Morrow J, Wu A, Roberts LJ II, Dettbarn WD. Diisopropyl phosphorofiurdate induced muscle hyperactivity associated with enhanced lipid peroxidation in vivo. Biochem Pharmacol 1996; 52: 357.
- Robineau P, Leclerg Y, Garbi A, Berrebi-Bertrand I, Lolievere LG. An organophosphorus compound Vx selectively inhibits the rat cardiac Na<sup>+</sup>-K<sup>+</sup>-ATPase isoform. FEBS Lett 1991; 28: 145.
- Basha PM, Nayeemunnisa. Effect of methylparathion on Na<sup>+</sup>-K<sup>+</sup>-and Mg<sup>+</sup>2-adenosine triphosphatase activity in developing central nervous system in rats. *Indian J Exp BioI* 1993; 31: 785-797.
- Blasiak J. Inhibition of (Na<sup>++</sup>K<sup>+</sup>)-ATPase by organophosphorus insecticides. *Polish J Environ Studies* 1995; 4: 23.
- Blasiak J. Allosteric inhibition of (Na<sup>++</sup>K<sup>+</sup>) ATPase by parathion and methyl parathion. *Pesticide Biochem Physiol* 1996; 54: 40-46.
- 25. Blasiak J. Inhibition of erythrocyte membrane (Ca<sup>+2</sup>-Mg<sup>+2</sup>) ATPase by organophosphorus/ insecticides parathion and methyl parathion. Comp Biochem Physiol Pharmacol Toxicol Endocrinol 1995; 110C: 119.
- 26. Hazarika A, Sarkar SN, Kataria M. Subacute toxicity of anilofos, a new organophosphorus herbicide in male rats: Effect on lipid peroxidation and ATPase activity. *Indian J Exp Biol* 2001; 39: 1113-1117.